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Department of Zoology, Madras Christian College, Chennai, Tamil Nadu, India Laboratory evaluation on the efficacy of *Bacillus thuringiensis* var. *israelensis* Berliner 1915 (Bacillaceae) on the instars of the dengue vector *Aedes aegypti* Linnaeus 1762 (Diptera: Culicidae) and against the adults of *Diplonychus indicus* Venk. & Rao (Hemiptera: Belostomatidae) as non-target organism

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#### **Abstract**

Mosquito control agents based on bacterial insecticides are the most widely used microbial pesticides as they are environment friendly alternatives to chemical insecticides and hold tremendous potential with improved efficacy. On the other hand, it is also necessary to preserve the non-target organisms, especially those that predate upon target mosquito larvae. Since, chemical larvicides are largely non-selective owing to their deleterious role on non-target organisms, bacterial insecticides when used do not cause harm to them. Therefore, in the present investigation, the toxicity of *Bacillus thuringiensis* var. *israelensis* was evaluated on all the larval instars of *Aedes aegypti* at concentrations of 0.1, 0.2, 0.3 and 0.4%. The LC<sub>50</sub> values after 24, 48 and 72 hours against the first, second, third and fourth instars were 0.20, 0.19 and 0.17%; 0.16, 0.15 and 0.15%; 0.19, 0.18 and 0.16%; 0.14, 0.12 and 0.12% respectively. Further, the adults of *Diplonychus indicus* which were tested as the non-target organism was exposed to the same concentrations of bacterial formulation and the results indicated that they did not have toxic effect and that the treated adult bugs were normal with high predatory potential. The present research indicated that integration with appropriate biocides may not interfere with the biocontrol potential of the predator in controlling mosquitoes.

Keywords: Bacillus thuringiensis var. israelensis, Aedes aegypti, larvicidal, Diplonychus indicus

#### Introduction

The most important event in the 20th century in the history of vector control was the discovery of mosquitocidal strains of bacterial insecticides. Mosquito control agents based on bacterial insecticides are the most widely used group of microbial pesticides as they are environment friendly alternatives to chemical insecticides [1]. Considerable research has been conducted on bacterial insecticides over the last decades, and major successes have been obtained. The significant advantages of bacterial insecticides over chemical insecticides have been responsible for their fast introduction into large-scale routine operations for mosquito control [2]. Bacillus species are potent pathogens of mosquitoes [3-5] with increased toxicity against its larval stage [6], as they hold tremendous potential with improved efficacy since they produce parasporal crystal proteins (endotoxin) during sporulation which are considered to possess mosquitocidal activity against the larval populations of Aedes, Anopheles and Culex species [7-9]. On the other hand, it is also necessary to preserve the diversity of organisms, especially those competitors or predators upon target mosquito larvae; and bacterial insecticides when used do not cause harm to the non-target organisms. Therefore, the present investigation was carried out to evaluate the toxicity of *Bacillus thuringiensis* var. israelensis on the larval instars of the dengue vector, Aedes aegypti, and against the adults of Diplonychus indicus as the non-target organism.

## 2. Materials and Methods

# 2.1. Microbial collection and preparation

*Bacillus thuringiensis* var. *israelensis* was obtained from Inbiotics laboratory, Nagercoil, Kanyakumari district, Tamil Nadu, India. The required quantity of this microbe was thoroughly mixed with distilled water for bioassay.

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#### 2.2. Collection of Diplonychus indicus

Diplonychus indicus adults collected from a nearby pond at Nagercoil, Kanyakumari district, Tamil Nadu, India were brought to the laboratory, maintained in a glass aquaria and were fed with mosquito larvae. The eggs were allowed to hatch into the instars and the adults were reared.

## 2.3. Larvicidal bioassay

Larvicidal bioassay was carried out as per the guidelines of World Health Organization with minor modifications [10]. Eggs strips of F<sub>1</sub> generation of laboratory colonized Aedes aegynti mosquitoes obtained from Entomology Research Institute, Lovola College, Chennai, Tamil Nadu, India were allowed to hatch. The study was conducted by introducing 25 first instar larvae into glass beakers (500mL) containing formulations of Bacillus thuringiensis var. israelensis (0.1, 0.2, 0.3 and 0.4%) each. The same procedure was performed for the second, third and fourth instars. Distilled water as control was maintained separately and run simultaneously. Mortality was observed 24, 48 and 72 hours after treatment. A total of six replicates per trial and a total of three trials for each concentration were carried out. Moribund larvae were scored dead when they showed no signs of movement when probed by a needle at their respiratory siphon.

# 2.4. Effect of *Bacillus thuringiensis israelensis* on non-target organism

Diplonychus indicus adults were selected as non-target organism and were acclimatized to the laboratory condition, where they were kept in an environment similar to their natural habitat. Five adults were exposed to the same concentrations of the bacterial formulation kept in a glass trough (1L). Distilled water without the bacterial formulation served as control was maintained separately and run parallel. Numbers of dead adults were recorded after 24, 48 and 72 hours of exposures and percentage mortality was

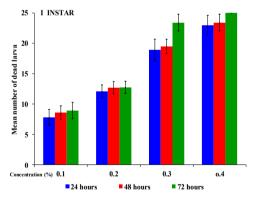
calculated. A total of six replicates per trial and a total of three trials for each concentration were carried out.

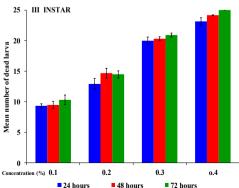
# 2.5. Statistical analysis

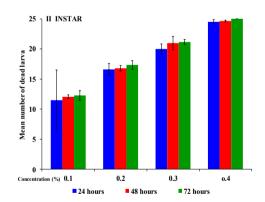
The per cent larval mortality was calculated and corrections for control mortality (5-20%) if necessary was done using Abbott's formula [11]. Statistical analysis of all mortality data were subjected to probit analysis. Chi-square and regression analysis tests were used to determine if the mortality in treated bioassays significantly differed from that of the controls and at which doses in particular and the differences were considered as significant at P=0.05 level. All statistics were conducted in IBM SPSS Statistics v22 with significance set at 95% confidence [12].

#### 3. Results

No larval mortality was observed in control. The mean and percentage mortality of Aedes aegypti larval instars treated with the Bacillus thuringiensis var. israelensis after 24, 48 and 72 hours are presented in Figure 1 and Table 1. One hundred per cent larval mortality was achieved at the highest concentration at 72 hours against the first, second and third instars and at 24 hours against the fourth instar. The LC<sub>50</sub> values after 24, 48 and 72 hours against the first, second, third and fourth instars were 0.20, 0.19 and 0.17%; 0.16, 0.15 and 0.15%; 0.19, 0.18 and 0.16%; 0.14, 0.12 and 0.12% respectively. Chi-square analysis revealed the data to be significant at P=0.05 level. Regression analysis revealed that mortality rates were positively correlated with the periods of exposure and R<sup>2</sup> values neared the value of 1. The same concentrations of *Bacillus thuringiensis* var. israelensis formulation when tested against the non-target organism did not have toxic effect even after 72 hours. The treated adults of Diplonychus indicus were normal and the predatory potential was high.







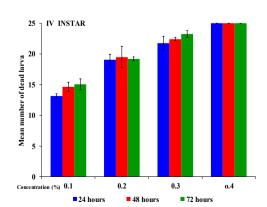


Fig 1: Mean mortality of Aedes aegypti larval instars on exposure to Bacillus thuringiensis israelensis

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 Table 1. Statistical inferences on the mortality of Aedes aegypti larval instars on exposure to Bacillus thuringiensis israelensis

Hours	Concentration (%)				LC <sub>50</sub> (%)	95% Confidence Limit		LC <sub>90</sub> (%)	95% Confidence		Slope ±SE	Intercept ±SE	χ²
									Limit				
	0.1	0.2	0.3	0.4		LL	UL	(70)	LL	UL			
I instar													
24	$31.22 \pm 1.32$	$48.33 \pm 1.08$	$75.65 \pm 1.78$	$91.92 \pm 1.64$	0.20	0.16	0.24	0.37	0.32	0.45	$7.63 \pm 1.15$	$-1.55 \pm 0.27$	3.34*
48	$34.32 \pm 1.12$	$50.80 \pm 1.02$	$78.00 \pm 1.18$	$93.64 \pm 1.38$	0.19	0.15	0.22	0.36	0.31	0.43	$7.62 \pm 1.14$	$-1.46 \pm 0.26$	4.10*
72	$35.66 \pm 1.33$	51.00 ±1.00	93.64 ±1.38	$100.00 \pm 0.00$	0.17	0.14	0.19	0.29	0.25	0.35	10.52 ±1.59	-1.78 ±0.31	4.66*
II instar													
24	$46.00 \pm 5.00$	$66.32 \pm 0.98$	$80.00 \pm 0.82$	98.00 ±0.38	0.16	0.03	0.25	0.32	0.23	0.66	$7.95 \pm 1.20$	-1.30 ±0.25	6.08*
48	48.23 ±0.32	67.21 ±0.45	83.83 ±1.12	98.64 ±1.56	0.15	0.02	0.25	0.31	0.22	0.67	$8.09 \pm 1.22$	-1.25 ±0.25	6.38*
72	49.00 ±0.83	$69.32 \pm 0.72$	84.62 ±0.40	$100.00 \pm 0.00$	0.15	0.01	0.25	0.29	0.20	0.69	$9.06 \pm 1.37$	-1.37 ±0.27	7.11*
III instar													
24	$37.33 \pm 0.33$	$51.66 \pm 0.88$	$80.00 \pm 0.58$	$92.66 \pm 0.60$	0.19	0.15	0.22	0.36	0.31	0.43	$7.62 \pm 1.14$	$-1.46 \pm 0.26$	4.10*
48	$38.00 \pm 0.57$	58.72 ±0.80	81.30 ±0.32	96.64 ±0.88	0.18	0.14	0.21	0.33	0.29	0.40	$8.29 \pm 1.23$	-1.49 ±0.27	3.74*
72	41.33 ±0.80	$58.00 \pm 0.58$	83.64 ±0.32	$100.00 \pm 0.00$	0.16	0.13	0.19	0.30	0.26	0.36	$9.33 \pm 1.39$	-1.55 ±0.28	5.04*
IV instar													
24	52.66 ±0.36	$76.33 \pm 0.88$	87.00 ±1.15	$100.00 \pm 0.00$	0.14	-0.08	0.26	0.28	0.18	0.99	$9.10 \pm 1.39$	-1.27 ±0.26	9.06*
48	$58.60 \pm 0.76$	$78.00 \pm 1.76$	89.66 ±0.26	$100.00 \pm 0.00$	0.12	-0.20	0.27	0.26	0.17	1.36	$9.36 \pm 1.46$	-1.20 ±0.26	9.90*
72	$60.32 \pm 0.88$	$76.82 \pm 0.32$	93.00 ±0.56	$100.00 \pm 0.00$	0.12	-0.07	0.23	0.25	0.16	0.83	10.15 ±1.59	$-1.26 \pm 0.27$	8.33*

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#### 4. Discussion

Bacillus thuringiensis var. israelensis are ideal biological control agents against mosquitoes and blackflies due to their efficacy and relative specificity [13, 14]. The efficacy of Bacillus thuringiensis var. israelensis formulations has been demonstrated in a variety of habitats against a multitude of mosquito species. Since the discovery of Bacillus thuringiensis serovariety israelensis [15, 16], formulations of these bacteria have become the predominant non-chemical means employed for control of mosquito larvae. The toxins of Bacillus thuringiensis var. israelensis and their mode of action, efficacy and factors that affect larvicidal activity. development of resistance, safety, and their roles in integrated mosquito control has opened a new way in mosquito control [17-20]. In the present study as the concentration of Bacillus thuringiensis var. israelensis increased the mortality rate of Aedes aegypti also increased. Bacillus thuringiensis var. israelensis is toxic to mosquitoes and their toxicity is commonly imputed to the parasporal endotoxins which are produced during sporulation time. These endotoxins are assimilated by the larvae to accomplish toxicity. Bacillus thuringiensis var. israelensis also produce different insecticidal crystal (endotoxins), and their toxicity has been determined [21, 22]. These toxins, when ingested by the larvae, damage the gut tissues, leading to gut paralysis, thereafter the infected larvae stop feeding and finally die due to the combined effects of starvation and impairment of midgut epithelium [23, 24]. Nevertheless, biotic and abiotic factors influence the larvicidal activity of Bacillus thuringiensis var. israelensis based on mosquito species, their respective feeding strategies, rate of ingestion, age and density of larvae. habitat factors (temperature, depth of water, turbidity). formulation factors (type of formulation, toxin content, how effectively the material reaches the target, and settling rate), storage conditions, means of application and frequency of treatments [25]. Amalraj et al. [26] reported that when it was applied for operational field applications the dipteran species were affected, but had no effect on Cladocerans. In the present study too, they caused toxicity to Aedes aegypti larvae and were not toxic to the instars and adults of Diplonychus indicus. Russel et al. [27] reported that water dispensable Bacillus thuringiensis var. israelensis formulation were toxic to the third instars of Aedes aegypti, Culex sittens, Culex annulristris and Culex quinquefasciatus but had no effect on the adults of Diplonychus indicus and the same trend was observed in the present study also. Nonetheless, Boisvert and Boisvert [28] reviewed the effect of Bacillus thuringiensis israelensis on target and non-target organisms and stated that, general predictions about the effect of Bacillus thuringiensis israelensis on non-target organisms may be difficult to make due to differences in (i) species tested, (ii) laboratory conditions and field methodology, and (iii) different formulations used.

# 5. Conclusion

The present research indicated that *Diplonychus indicus*, a good biocontrol agent especially on mosquito larva is not affected by biocides like *Bacillus thuringiensis* var. *israelensis*. This large safety margin of this bacterial insecticide for non-target organisms indicates their suitability for mosquito control programmes in areas where protection of the natural ecosystem is important. Therefore, its integration with appropriate biocides may not interfere

with the biocontrol potential of the predator in controlling mosquitoes.

# 6. References

- De Barjac H. Characterization and prospective view of Bacillus thuringiensis subsp israelensis. In: Bacterial control of mosquitoes and black flies. (Eds.) de Barjac H, Sutherland DJ. Rutgers University Press, New Jersey. 1990, 10-15.
- 2. Becker N. The use of *Bacillus thuringiensis* subsp. *israelensis* (Bti) against mosquitoes, with special emphasis on the ecological impact. Israel Journal of Entomology. 1998; 32:63-69.
- 3. Porter AG, Davidson E, Liu JW. Mosquitocidal toxins of bacilli and their genetic manipulation for effective biological control of mosquitoes. Microbiological Reviews. 1993; 57:838-861.
- 4. Cooping LG, Menn JJ. Biopesticides: a review of their action, applications and efficacy. Pest Management Science. 2001; 56:651-676.
- 5. Wirth MC, Delecluse A, Walton WE. Laboratory selection for resistance to *Bacillus thuringiensis* subsp. *jegathesan* or a component toxin, Cry 11B, in *Culex quinquefasciatus* (Diptera: Culicidae). Journal of Medical Entomology. 2004; 41(3):435-441.
- Goldberg LH, Margalit J. A bacterial spore demonstrating rapid larvicidal activity against Anopheles sergentii, Uranotaenia unguiculata, Culex univittatus, Aedes aegypti and Culex pipiens. Mosquito News. 1977; 37:355-358.
- 7. Federici BA, Park HW, Sakano Y. Insecticidal protein crystals of *Bacillus thuringiensis*. In: Inclusions in prokaryotes. (Ed.) Shively JM., Springer-Verlag, Berlin, Heidelberg, Germany. 2006, 195-236.
- 8. Park HW, Mangum CM, Zhong H, Sabrina SR. Isolation of *Bacillus sphaericus* with improved efficacy against *Culex quinquefasciatus*. Journal of the American Mosquito Control Association. 2007; 23:478-480
- Revathi K, Chandrasekaran R, Thanigaivel A, Kirubakaran SA, Narayanan SS, Nathan SS. Effects of Bacillus subtilis metabolites on larval Aedes aegypti L. Pesticide Biochemistry and Physiology. 2013; 107:369-376
- WHO. Guidelines for laboratory and field testing of mosquito larvicides. WHO, Geneva, WHO/CDS/WHOPES/GCDPP/13. 2005.
- 11. Abbott WS. A method of computing the effectiveness of an insecticide. Journal of Economic Entomology. 1925; 18:265-267.
- 12. SPSS. IBM SPSS Statistics for Windows, Version 22.0. Armonk, NY: IBM Corp. 2010.
- 13. Sharma SN, Sukla RP, Mittal PK, Adak T, Kumar A. Efficacy of a new formulation of *Bacillus thuringiensis* var *israelensis* (Bti) in laboratory and field conditions of Kumaun foothills of Uttaranchal, India. Journal of Communicable Diseases. 2003, 290-299.
- 14. Hongya Z, Changin Y, Jingye H, Lin L. Susceptibility of field populations of *Anopheles sinensis* (Diptera: Culicidae) to *Bacillus thuringiensis* subsp. *israelensis*. Biocontrol Science and Technology. 2004; 14:321-325.
- 15. Ali A, Chowdhury MA, Aslam AFM, Ameen M, Hossain MI, Habiba DB. Field trails with *Bacillus sphaericus* and *Bacillus thuringiensis* serovar.

Acta Entomology and Zoology http://www.actajournal.com

israelensis commercial formulations against *Culex quinquefasciatus* larvae in suburban Dhaka, Bangladesh. Medical Entomology and Zoology. 2000; 51:257-264.

- 16. Fullinger U, Knols BGJ, Becker N. Efficacy and efficiency of new *Bacillus thuringiensis* var. *israelensis* and *Bacillus sphaericus* formulations against Afrotropical anophelines in Western Kenya. Tropical Medicine and International Health. 2003; 8(1):37-47.
- 17. Dennet JA, Lampman RL, Novak RJ, Meisch MV. Evaluation of methylated soy oil and water based formulations of *Bacillus thuringiensis* var. *israelensis* and golden bear oil (GB- 1111) against *Anopheles quadrimaculatus* larvae in small rice plots. Journal of the American Mosquito Control Association. 2000; 16(4):342-345.
- 18. Chansang U, Bhumiratana A, Kittayapong P. Combination of *Mesocyclops thermocyclopoides* and *Bacillus thuringiensis* var *israelensis*: A better approach for the control of *Aedes aegypti* larvae in water containers. Journal of Vector Ecology. 2004; 29:218-226.
- 19. Zahiri NS, Federic BA, Mulla MS. Laboratory and simulated field evaluation of a new recombinant of *Bacillus thuringiensis* ssp. *israelensis* and *Bacillus sphaericus* against *Culex* mosquito larvae (Diptera: Culicidae). Journal of Medical Entomology. 2004; 41(3):423-429.
- 20. Zahiri NS, Mulla MS. Non-larvicidal effects of *Bacillus thuringiensis israelensis* and *Bacillus sphaericus* on oviposition and adult mortality of *Culex quinquefasciatus* Say (Diptera: Culicidae). Journal of Vector Ecology. 2009; 30(1):155-162.
- 21. Chilcott CN, Kalmakoff J, Pillai JS. Characterization of proteolytic activity associated with *Bacillus thuringiensis* var. *israelensis* crystals. FEMS Microbiological Letters. 1983; 18:37-41.
- Russel TL, Brown MD, Purdie DM, Ryan PA, Kas BH. Efficacy of Vectobac (*Bacillus thuringiensis* variety *israelensis*) formulations for mosquito control in Australia. Journal of Economic Entomology. 2003; 96(6):1786-1791.
- 23. Aronson AI, Shai Y. Why *Bacillus thuringiensis* insecticidal toxins are so effective: unique features of their mode of action. FEMS Microbiological Letters. 2001; 195:1-8.
- 24. Betz FS, Hammond BG, Fuchs RL. Safety and advantages of *Bacillus thuringiensis* protected plants to control insect pests. Regulatory Toxicology and Pharmacology. 2000; 32:156-173.
- 25. Zhu YC, Kramer KJ, Oppert B, Dowdy AK. cDNAs of aminopeptidase-like protein genes from *Plodia interpunctella* strains with different susceptibilities to *Bacillus thuringiensis* toxins. Insect Biochemistry and Molecular Biology. 2000; 30:215-224.
- 26. Furnani SC, Arita, Tsutsuni L. Use of *Bacillus thuringiensis israelensis* and methoprene to control the Asian tiger mosquito, *Aedes albopictus* (Skuse) (Diptera: Culicidae) larvae and pupae populations in non-circulating hydroponic tanks. Hawaiian Entomological Society. 2002; 35:113-119.
- Amalraj DD, Sahu SS, Jambulingam P, Doss PPS, Kalyanasundaram M, Das PK. Efficacy of aqueous suspension and granular formulations of *Bacillus*

- thuringiensis (VectoBac) against mosquito vectors. Acta Tropica. 2000; 25:243-246.
- 28. Boisvert M, Boisvert J. Effect of *Bacillus thuringiensis* var. *israelensis* on target and non-target organisms: A review of laboratory and field experiments. Biocontrol Science and Technology. 2000; 10:517-561.